

1 **Wetting and drying cycles, organic amendments, and gypsum play a key role in structure formation and**
2 **stability of sodic Vertisols**

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13 **Abstract**

14 In the natural environment, soils undergo wetting and drying (WD) cycles due to precipitation
15 and evapotranspiration. The WD cycles have a profound impact on soil physical, chemical, and
16 biological properties and drive the development of structure in soils. Degraded soils are often
17 lacking structure and the effect of organic amendments and WD cycles on structure formation
18 of these soils is poorly understood. The aim of this study was to evaluate the role of biotic and
19 abiotic factors on aggregate formation and stabilisation of sodic soils after the addition of
20 gypsum and organic amendments (feedlot manure, chicken manure, lucerne pellets, and
21 anionic poly acrylamide). Amended soils were incubated at 25°C over four WD cycles, with
22 assessment of soil microbial respiration, electrical conductivity, pH, sodium adsorption ratio
23 (SAR), aggregate stability in water (ASWAT), aggregate size distribution, and mean weight
24 diameter. Our results demonstrate that WD cycles can improve aggregate stability after the
25 addition of amendments in sodic Vertisols, but this process depends on the type of organic
26 amendment. Lucerne pellets resulted in highest soil microbial respiration, proportions of large
27 macroaggregates (>2000 µm), and mean weight diameter. In contrast, dispersion was

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29 significantly reduced when soils were treated with chicken manure, whilst [anionic](#)
30 [polyacrylamide](#), only had a transient effect on aggregate stability. When these organic
31 amendments were applied together with gypsum, the stability of aggregates was further
32 enhanced, and dispersion became negligible after the second WD cycle. The formation and
33 stability of small macroaggregates ([2000-250 µm](#)), was less dependent on the type of organic
34 amendments and more dependent on WD cycles as the proportion of small macroaggregates
35 also increased in control soils after four WD cycles, highlighting the role of WD cycles as one
36 of the key factors that improves aggregation and stability of sodic Vertisols.

37 **Key words:** Sodic soils, organic amendments, aggregate stability, mean weight diameter,
38 wetting and drying cycles

39 1 Introduction

40 Soils are subjected to seasonal and daily variations of water and temperature. These variations
41 effect the physical, biological, and chemical properties of soils. Natural variations in soil water
42 content can lead to wetting and drying (WD) cycles, which are affected by rainfall, solar
43 radiation, capillarity, wind, [condensation \(Utomo and Dexter 1982\)](#) and evapotranspiration. In
44 most terrestrial ecosystems, surface soils experience rapid changes in soil water content with a
45 longer dry period followed by a relatively rapid re-wetting (Borken and Matzner 2009).
46 Southern Queensland (Australia) usually has hot wet summers with cool dry winters, and soils
47 of this region and other semi-arid areas are particularly susceptible to drying and re-wetting
48 stresses due to the infrequency of rainfall. In the field, most soils experience more than one
49 WD cycle throughout the year. These WD cycles have a substantial influence on soil
50 aggregation and structure stabilisation because of its direct effect on hydration of minerals, and
51 indirect effect on [plant ecology and](#) soil microbial activity (Denef *et al.* 2001; Cosentino *et al.*
52 2006).

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56 Soil aggregation is an important mechanism for stabilisation of soil organic matter (Six *et al.*
57 2000a). Furthermore, it also supports soil fertility as it reduces soil erosion and controls soil
58 aeration, water infiltration, hydraulic conductivity and nutrient cycling (Oades 1984; Six *et al.*
59 2000b). Soil aggregation is caused by various aggregate stabilising compounds, which work
60 together at different spatial scales (Tisdall and Oades 1982). Aggregate formation also depends
61 on soil microbial activity, since the latter, influences the production of binding materials such
62 as microbial exudates and hyphae (Rahman *et al.* 2017). Aggregate stability often exhibits
63 seasonal and inter-annual variability that is controlled mainly by rainfall, temperature,
64 humidity, insolation, and organic matter (Perfect *et al.* 1990).

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65 Sodic soils are generally characterised as soils with poor structure which makes those soils
66 difficult to work when wet or dry (Rengasamy and Olsson 1991). Poor structural stability of
67 sodic soils restricts seedling emergence and root growth which directly limits crop growth and
68 development, and indirectly affects plant nutrition by limiting water infiltration, nutrient
69 uptake, and gaseous exchange (Curtin and Naidu 1998). Rehabilitation of sodic soils requires
70 an understanding of the complex interactions between biological and physico-chemical factors
71 that contributes to soil structure formation and stability (Oades 1993; Nelson and Oades 1998).

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72 Traditionally the management practices used to improve the structure of sodic soils involves
73 the displacement of Na ions from the soil exchange complex with the help of divalent cations
74 such as Ca or increasing the ionic strength of soil solution (Ghosh *et al.* 2010), both of which
75 can be achieved by the application of gypsum to these soils. The effect of gypsum on increasing
76 ionic strength is immediate, but short lived. In contrast, the effect of gypsum for providing the
77 counter ion (Ca to replace Na) is permanent unless additional Na is added to system (e.g., using
78 poor quality irrigation water). Another frequently used management practice to ameliorate
79 sodic soils is the use of organic amendments. Addition of organic matter effect aggregate
80 stability within a period of days to weeks due to the stimulation of microbial activity (Six *et al.*

83 2004), depending upon the quality and quantity of organic matter (Monnier, 1965 cited in
84 Abiven *et al.* (2009)). While organic matter increase soil microbial respiration resulting in the
85 formation of extracellular polysaccharides which help in the formation of soil aggregates
86 (Bossuyt *et al.*, 2001), studies investigating the effect of organic amendments in improving the
87 soil structure are inconclusive. For instance, the extracellular polysaccharides and large
88 polyanions can bind clay particles into stable macroaggregates. On the other hand, organic
89 anions can enhance dispersion by increasing the negative charge on clay particles and by
90 complexing calcium and other polyvalent cations (such as those of aluminium), hence reducing
91 their activity in soil solution (Ghosh *et al.* 2010)
92 „Apart from the changes in soil structure due to the addition of different ameliorants, WD cycles*
93 can lead to more intensive changes in structure of soils dominated by smectitic clays
94 (Vertisols), through physical processes (Utomo and Dexter 1982; Deneff *et al.* 2001). These
95 soils are generally characterised as self-mulching soils as they exhibit shrink-swell properties
96 imposed by the WD cycles (Pal *et al.*, 2012). Vertisols cover a total of an estimated 340 million
97 ha in the world (Australia, Asia, Africa, and America), out of which approximately 150 million
98 ha is potential crop land. However, the physical properties and moisture regime of Vertisols
99 represents serious management constraints (Pal *et al.* 2012). Sodic Vertisols are common in
100 arid parts of the world. The effect of sodicity on the physical properties of Vertisols is still a
101 subject of debate. For example, Rahman *et al.* (2018) reported that WD cycles improved
102 (increased) the mean weight diameter (MWD, a proxy of aggregate stability measurement) of
103 Vertisols when treated with maize straw after four WD cycles. In a similar manner, WD cycles
104 also increased the proportion of large macro aggregates of smectite Vertisols (Bravo-Garza *et*
105 *al.* 2009). Peng *et al.* (2011), comparing swelling and non-swelling soils, reported that WD
106 cycles decreased the MWD of swelling soils but not of non-swelling soils. However, Six *et al.*
107 (2000b) found that repeated WD cycles decreased the MWD due to physical disturbance of

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121 [aggregates](#), with this being related to the loss of soil organic matter. These apparently
122 contradictory results can potentially be explained on the basis of the initial conditions of the
123 soil, such as the physical conditions of aggregates, organic matter content and quality, and the
124 intensities and duration of the WD cycles (Cosentino *et al.* 2006).

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125 ~~Similarly, studies investigating the effects of WD cycles on microbial activity were~~
126 inconclusive, with results differing due to varying experimental designs, incubation period and
127 temperatures, soil properties, and treatments applied. Xiang *et al.* (2008) found that multiple
128 WD cycles increased the microbial respiration of grassland soils up to 6-fold when compared
129 to the soil that remained wet. Drying and rewetting of these soils gave a new pulse of respiration
130 with each WD cycle, but when these soils were kept at constant water content, microbial
131 respiration decreased to almost zero. [An increase in cumulative respiration from forest soils of](#)
132 [China was also observed when these soils experience WD cycle compared to the constant](#)
133 [moisture conditions \(Zhang *et al.* 2022\).](#) In contrast, however, Rahman *et al.* (2018) reported
134 that repeated WD cycles in Vertisols decreased soil respiration significantly but that the
135 magnitude of this decrease became smaller over the time. [Similarly, Yu *et al.* \(2014\) found that](#)
136 [repeated WD cycles decreased the cumulative soil respiration compared to constant moist](#)
137 [conditions in a loamy sand soil.](#)

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138 Although the interaction of biotic and abiotic factors and its effect on aggregate stability and
139 formation is complex and inconsistent, little effort has been put into studying the underlying
140 mechanisms, particularly in sodic Vertisols. Furthermore, there is little information available
141 on the relationship between aggregate formation and stability following repeated WD cycles
142 after addition of gypsum and organic amendments. Thus, in the present study we used two
143 sodic Vertisols, [with the aim to: i\) determine the role of gypsum and different organic](#)
144 [amendments on aggregate formation and stability, ii\) explore the combined effect of gypsum](#)
145 [and organic amendments on soil physico-chemical and microbial properties, iii\) investigate the](#)

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153 [effect of WD cycles on microbial respiration](#), iv) [assess the effects of WD cycles on aggregate](#)
154 [formation and stability](#), and v) [determine how many WD cycles are needed to improve](#)
155 [aggregate stability](#). We hypothesised that i) [organic amendments will increase the microbial](#)
156 [respiration and improves the formation of large macroaggregates and MWD](#), (ii) [gypsum will](#)
157 [improve aggregate stability due to increases Ca concentration and ionic strength](#), (iii) [organic](#)
158 [amendments act synergistically with gypsum on aggregation](#), and iv) [repeated WD cycles will](#)
159 [increase the process of aggregate formation and stability](#).

160 2 **Materials and methods**

161 2.1 **Soils**

162 The two soils used in this experiment were collected from a farm located in southern
163 Queensland near Goondiwindi (28.54° S, 150.30° E), Australia. The soils were being cropped
164 using a maize (*Zea mays*) and wheat (*Triticum aestivum*) rotation system. Both soils were
165 classified as sodic Vertisols according to the FAO-World Reference Base (2015) (Vertisols in
166 the Australian Soil Classification). Soils were collected [with a shovel](#) to a depth of 10 cm at
167 two locations from the cropped land. These sites were selected as they were in close proximity
168 to each other but were slightly different in regard to their physical [properties](#) (Table 1) with
169 Soil 1 being more dispersive than Soil 2. [Three core samples were also collected for the](#)
170 [measurement of](#) bulk density [from each site \(Soil 1 and Soil 2\)](#) and dried at 105°C for several
171 days. After collection, soils were air dried, and the larger clods gently broken into smaller
172 aggregates by hand. The soils were then passed through a 10 mm sieve to remove stones, visible
173 roots, and plant litter. For chemical analysis, a portion of each soil was sieved to <2 mm. Soil
174 pH (ISO, 2005) and electrical conductivity (EC) (ISO, 1994) were measured in 1:5 soil: water
175 suspension. [Particle size analysis was performed using the pipette method \(Day, 1965\)](#), and
176 field water capacity (-10 kPa) was measured using a pressure plate apparatus (Cassel and
177 Nielsen, 1986). Exchangeable cations (Ca²⁺, Mg²⁺, Na⁺, K⁺), and effective cation exchange

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184 capacity were determined after pre-washing with 60% ethanol, and then leaching with non-
 185 alcoholic 1 M NH₄Cl solution at pH 7 (Tucker, 1985). The exchangeable sodium percentage
 186 was calculated as exchangeable Na concentration divided by the effective cation exchange
 187 capacity. Total organic C, and total N analyses were conducted using a LECO TruMac
 188 instrument using the Dumas method (Nelson and Sommers, 1996). The electrochemical
 189 stability index was used as an indication of the potential for soil dispersion. The threshold
 190 electrochemical stability index below which structural breakdown occurs is 0.05 (McKenzie,
 191 1998). The dispersion index (DI), another measure of soil structural stability, is defined as the
 192 amount of dispersed silt+clay expressed as a percentage of the total silt+clay of the soil (see
 193 section 2.3) (Mustafa and Letey, 1969). Soil dispersion assessed by aggregate stability in water
 194 (ASWAT) is described in detail in Section 2.3.

195 **Table 1** *Physico-chemical properties of soil samples*

	Soil 1	Soil 2
197 pH (1:5 water)	7.07	7.03
198 EC (1:5 dS m ⁻¹)	0.26	0.22
199 Sand (%)	34	37
200 Silt (%)	17	17
201 Clay (%)	49	46
202 Bulk density (g cm ⁻³)	1.29	1.29
203 Field capacity (%)	33	29
204 Total organic C (g kg ⁻¹)	8.8	9.8
205 Total N (g kg ⁻¹)	0.90	0.90
Effective cation exchange capacity (cmol ⁽⁺⁾ kg ⁻¹)	23	24
Exchangeable sodium percentage	15	16
Electrochemical stability index	0.02	0.01
ASWAT	15	11
DI (%)	48	28

206 **2.2 Collection of different organic materials used as amendments**

207 Four different organic amendments were used for this study, viz. feedlot manure (FLM),
208 chicken manure (CM), lucerne pellets (LP), and polyacrylamide (PAM). The FLM, and CM
209 were collected from cattle, and chicken sheds, respectively, located at The University of
210 Queensland (Gatton, Australia) before being air dried, whilst the LP, was a commercial animal
211 feed product. The four organic amendments were chosen because of three reasons: 1) they were
212 easily available and are being used by farmers, 2) LP is used as green manure and studies have
213 shown it is effective in ameliorating sodic soils, and 3) PAM is used in mining and construction
214 to treat sodic dispersive soils. Furthermore, these amendments were different in terms of their
215 chemical properties (Table 2) and C functional groups (Niaz et al., 2022) and may give a good
216 contrast between the amendments. All organic amendments were ground and sieved through a
217 0.5 mm sieve in order to minimise the effect of different size particles and to create a
218 homogenised sample. Anionic PAM Flobond L33 liquid (SNF Australia) had a charge density
219 of 30% and the molecular weight was 12-15 million Dalton (as per product specifications).
220 Gypsum (CaSO₄.2H₂O) was laboratory grade obtained from Sigma Aldrich.

221 A chemical analysis of the organic amendments was performed before mixing with the soils
222 (Table 2). Total C and N analysis were performed using the LECO TruMac instrument with
223 the Dumas method (Nelson and Sommers, 1996). The major cations, including Na⁺, Ca²⁺,
224 Mg²⁺, and K⁺, were quantified by inductively coupled plasma-optical emission spectrometry
225 (ICP-OES) after nitric acid microwave digestion (Kovács et al., 1996). The pH and EC of
226 organic amendments were measured in (1:5) water suspensions after shaking for 1 h.

227 **Table 2** Chemical properties of organic amendments

	pH	EC (1:5)	TC	TN	C:N	Ca ²⁺	Na ⁺	Mg ²⁺	K ⁺
	(1:5)	(dS m ⁻¹)	(%)	(%)		(g kg ⁻¹)			
FLM	8.4	3.0	14.9	1.4	10:1	18.1	4.2	12.8	12.5

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CM	8.2	10.7	26.0	2.6	10:1	179.0	4.7	9.7	29.7
LP	5.6	5.7	43.8	3.1	14:1	13.8	2.9	3.7	12.8
PAM			49.7	0.9	56:1				

230 2.3 Incubation experiment

231 An incubation experiment was conducted using a complete randomised design. The treatments
 232 consisted of the five amendments (gypsum (G), PAM, FLM, CM, and LP) plus an unamended
 233 control. In addition, the FLM, CM, LP, and PAM were also applied in combination with G in
 234 order to check for synergistic effects between gypsum and organic amendments. This yielded
 235 a total of 10 treatments, each with three replicates, being a total of 60 experimental units. Soil
 236 samples (300 g) were mixed with the respective organic amendments (Table 3) added at rates
 237 that were commercially feasible (10 Mg ha⁻¹) except PAM which was added at 1 kg ha⁻¹. The
 238 gypsum requirement of both soil samples was calculated based on the formula given by Oster
 239 and Jayawardane (1998) as follows:

240 $Gypsum\ requirement\ (GR) = 0.00086 \times F \times D \times \partial b \times (CEC) \times (ESP_i - ESP_f) \dots (1)$ *

241 Where F is exchanged efficiency of Ca-Na and for this case considered equal to 1, D is the
 242 depth of soil to be reclaimed (cm), ∂b is soil bulk density (g/cm³), CEC is cation exchange
 243 capacity (cmol⁺/kg), ESP_i is initial soil exchangeable sodium percentage, ESP_f is final or
 244 desired exchangeable sodium percentage. For simplicity, a single gypsum rate of 2.5 Mg/ha
 245 was selected for both soils.

246 The experiment was performed jars with 12 cm diameter and 15 cm height, and the soil was
 247 packed to a height of 3-5 cm. The soil water content at field capacity (-10 kPa) was ~ 0.30 g
 248 water g⁻¹ soil. The WD cycles were imposed as follows. First deionised water was added to the
 249 soils (0.30 g g⁻¹ for Soil 1 and 0.31 g g⁻¹ for Soil 2) at 25°C, and after 1 d, the lids were removed
 250 and soils were allowed to dry at 25°C during which the soil water content decreased to air-dry
 251 conditions (~0.1 g water g⁻¹ soil). The dry-down typically completed in 14 d. The WD regime

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256 was applied four times with the total duration of the experiment being 60 d. Although the WD
 257 regime might not entirely represent the field conditions due to lack of plant growth, it is
 258 representative of the mean temperature and the field water content.

259 **Table 3** *Treatments and their application rates*

	Treatments	Application rates
1	Control	No amendment
2	G	2.5 Mg ha ⁻¹
3	FLM	10 Mg ha ⁻¹
4	CM	10 Mg ha ⁻¹
5	LP	10 Mg ha ⁻¹
6	PAM	1 kg ha ⁻¹
7	G + FLM	2.5 Mg ha ⁻¹ + 10 Mg ha ⁻¹
8	G + CM	2.5 Mg ha ⁻¹ + 10 Mg ha ⁻¹
9	G + LP	2.5 Mg ha ⁻¹ + 10 Mg ha ⁻¹
10	G + PAM	2.5 Mg ha ⁻¹ + 1 kg ha ⁻¹

260 Soil solutions (~3-5 ml) were extracted using polyacrylonitrile hollow fibre samplers (Menzies
 261 and Guppy, 2000) embedded in the jars containing the treated soils. Soil solution was collected
 262 four times at the start of each WD cycle after the water was added to soils. Soil solution pH,
 263 EC, NH₄⁺-N, NO₃⁻-N, dissolved organic carbon (DOC), Na, Ca, Mg, and K were measured as
 264 follows: NH₄⁺-N by the phenate method (Rice et al., 2017), NO₃⁻-N by cadmium reduction
 265 (Rice et al., 2017) using a segmented flow analyser (SEAL AA3), DOC using a total organic
 266 carbon liquid analyser (Shimadzu, Japan), and the cations using ICP-OES. The
 267 sodium adsorption ratio (SAR) was calculated from the soil solution concentrations of Na⁺,
 268 Ca²⁺, and Mg²⁺.
 269 Soil dispersion was determined using the ASWAT test (Field *et al.* 1997) and DI (Mustafa and
 270 Letey 1969). Air dried aggregates from each sample after completion of each cycle were placed
 271 in a Petri dish filled with deionised water. A visual assessment of aggregate dispersion was

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273 made after 10 min and 2 h, assessing dispersion on a scale of 0-4. The scores were as follows:
274 0: no dispersion, 1: slight dispersion (slight milkiness adjacent to aggregates in water), 2:
275 moderate dispersion (obvious milkiness), 3: strong dispersion (considerable milkiness with
276 about half of the material dispersed in water), and 4: complete dispersion (leaving sand particles
277 in a cloud of dispersed clay). Dispersion scores that were determined after 10 min and 2 h were
278 added together and then added to 8, thus giving a range of values from 9-16. Aggregates which
279 were not dispersed after 2 h were remolded, submerged in water and dispersion reassessed
280 again after 10 min and 2 h. Scores were given from 0-4 for both times and added (giving values
281 from 0-8). Thus, aggregates were considered stable if they had a low ASWAT score, and the
282 higher the ASWAT score, the more unstable the aggregate.

283 For the measurement of DI, 15 g of air-dry soil (< 2 mm) was placed in a sedimentation cylinder
284 with an approximate volume of 1.2 L. Deionised water was added to the cylinder to bring the
285 volume to a 1 L. The suspension was then shaken for 30 min on an end-over-end shaker. The
286 suspension was then stirred with 10 strokes of a plunger. The temperature was noted, the
287 suspension was allowed to settle, and after an appropriate time (8 h) the percentage of clay was
288 measured with hydrometer. The measurements were done after the first, second, and fourth
289 WD cycles. Results were expressed as DI (Mustafa and Letey 1969):

290
$$DI(\%) = 100 \times \frac{\text{silt+clay (easily dispersed,\%)}}{\text{silt+clay (particle size analysis,\%)}}$$

(2)

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291 Aggregate size distribution was also determined to quantify changes in large and small
292 aggregates with WD cycles. Air dried soil samples (25 g) were broken gently to pass through
293 a 10 mm sieve and then slowly wetted up in water for 5 min (Hernandez *et al.* 2017). They
294 were then wet sieved for 5 min at 33 oscillations per minute (Cook *et al.* 1992) using a set of
295 sieves: 2000 µm, 250 µm and 53 µm (Kemper and Rosenau 1986). Three replications were
296 made of each sample. The water level was adjusted so that the aggregates on the upper sieve
297 were submerged in water at the highest point of the oscillation. The material retained on each

299 sieve and the fraction passing through after wet sieving was collected, dried at 40 °C, and
300 weighed. (Kemper and Rosenau 1986). Measurements were made after the first, second, and
301 fourth WD cycles. The >2000 µm aggregates hereafter are referred to as large
302 macroaggregates, 2000-250 µm as small macroaggregates, 250-53 µm as micro-aggregates
303 (MIC), and <53 µm as silt+clay fraction. The MWD was calculated from the aggregate
304 fractions obtained after wet sieving on each sieve size, as follows:

305
$$MWD = \sum_{i=1}^n x^i w^i \quad (3)$$

306 Where x^i is the mean diameter of each fraction and w^i is the proportion of the total sample
307 weight in the corresponding size fraction (Kemper and Rosenau 1986).

308 2.4 Soil microbial respiration

309 The biolability (susceptibility to microbial decomposition) of the organic amendments was
310 determined by measuring CO₂ release over a 60 d incubation period comprising four WD
311 cycles. Briefly, 50 g of each soil sample was added to 250 mL jars with the relevant
312 amendments (Table 3) and covered with lids having two 5 mm holes. All the treatments were
313 replicated three times and randomised. The soils were wetted to field capacity on the first day
314 of incubation using deionised water and incubated at 25 °C. After the first day of soil
315 incubation, the jars were left open to allow the soil to dry at 25 °C. For the microbial respiration
316 measurements, rubber tubes with Luer locks were inserted into the holes and connected to a
317 CO₂ analyser (WMA-4 CO₂ analyzer, John Morris USA). The lids were kept closed for 1 h
318 prior to the measurement being taken. The readings were then converted in g C-CO₂ kg⁻¹ of
319 soil d⁻¹. Cumulative CO₂ produced was calculated as the sum of the daily rate and the interval
320 days between the two measurements (by linear extrapolation) for the incubation period. The
321 measurements were taken daily for the first 7 d. Thereafter, there was no significant change in
322 microbial respiration, so the measurements were performed after 10 d and 15 d of each WD
323 cycle.

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326 2.5 Statistical analysis

327 Statistical analyses were performed using R x 64 3.3.3 statistical software (R Core Team,
328 2020). Histograms were plotted to check the normality of each parameter separately and found
329 that transformation of data was not necessary. Soil microbial respiration was recorded on a
330 daily basis hence respiration data were subjected to analysis of variance with days as the
331 repeated measures (mixed effect model). All the remaining parameters including EC, pH,
332 $\text{NH}_4\text{-N}$ and $\text{NO}_3\text{-N}$, SAR, ASWAT, DI, aggregate size distribution, and MWD were measured
333 after each WD cycle. Hence, these data were analysed using a two-way analysis of variance
334 (ANOVA) taking treatments and WD cycles as two factors. The ANOVA for each soil was
335 conducted separately using general linear model. Tukey's honest significant difference was
336 used for pairwise comparisons between treatment means. After checking the significance of
337 data, mean data were graphed using Sigma Plot v14.0 (Systat Software Inc., 2017). Principal
338 component analyses (PCA) were performed to identify the roles of microbial, chemical and
339 physical properties after the addition of different treatments in each WD cycle.

340 3 Results

341 3.1 Soil aggregation dynamics measured by aggregate size distribution and mean 342 weight diameter

343 A significant ($p < 0.001$) interaction between WD cycles and amendments was found for all the
344 aggregate sizes in both soils, except for the proportion of large macroaggregates and silt+clay
345 fraction in Soil 2 (Fig. 1), although the main effects of treatments and WD cycles were highly
346 significant ($p < 0.001$) in Soil 2. The finding of a significant interaction between amendments
347 and WD cycles indicates that although aggregate size distribution changed with the WD cycles,
348 the pattern of change depended on the amendment. Overall, the proportion of large
349 macroaggregates decreased after the first WD cycle and increased by the end of fourth WD
350 cycle. Among the various amendments, the LP and G+LP treatments had the highest proportion

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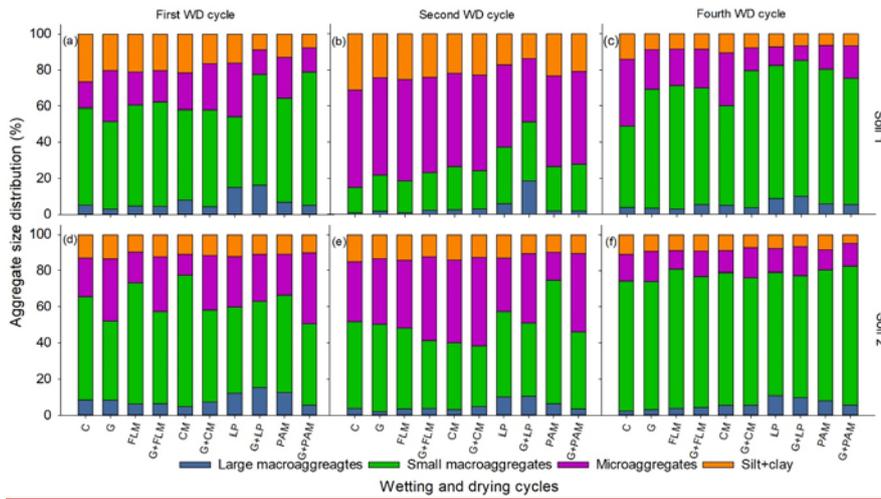
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358 of large macroaggregates, whereas PAM, FLM, and CM had little or no effect on large
359 macroaggregates for either soil. We also observed changes in the small macroaggregate
360 fraction, with this markedly decreasing by the second WD cycle but greatly increasing by the
361 fourth WD cycle. In contrast to the changes in small macroaggregates, the proportion of
362 microaggregates increased considerably (ca. two-fold) by the second WD cycle and greatly
363 decreased by the fourth WD cycle in both soils. The silt+clay fraction slightly increased by the
364 end of second WD cycle and again decreased slightly by the fourth WD cycle in both soils.
365 The MWD of both soils was calculated from the aggregate size distribution. The main effects
366 of amendments and WD cycles were found highly significant ($p < 0.001$) in both soils, however,
367 the interaction between amendments and WD cycles was significant only for Soil 1. It was
368 observed that MWD decreased after the second WD cycle (Fig. 2) and increased after the fourth
369 WD cycle, irrespective of amendments or soil, reflecting the changes in actual aggregate sizes
370 (see Fig.1). Expressing changes in MWD relative to the control or gypsum treated soil showed
371 that LP increased the MWD in both soils over the four WD cycles (Fig. S1), with PAM being
372 less effective than LP, whereas the other amendments had no significant effect.

373



374

375 *Figure 1. Distribution of the aggregate sizes in the two soils after addition of amendments after first,*
 376 *second, and fourth WD cycles: First WD cycle (a; Soil 1, and d; Soil 2), Second WD cycle (b; Soil 1,*
 377 *and e; Soil 2), and fourth WD cycle (c; Soil 1, and f; Soil 2) (C: control, G: gypsum, PAM: anionic*
 378 *polyacrylamide, FLM: feedlot manure, CM: chicken manure, LP: lucerne pellets). Large*
 379 *macroaggregates (>2000 μm); small macroaggregates (2000-250 μm); microaggregates (250-53 μm),*
 380 *and silt+clay fraction (<53 μm). Error bars have been omitted to improve readability.*

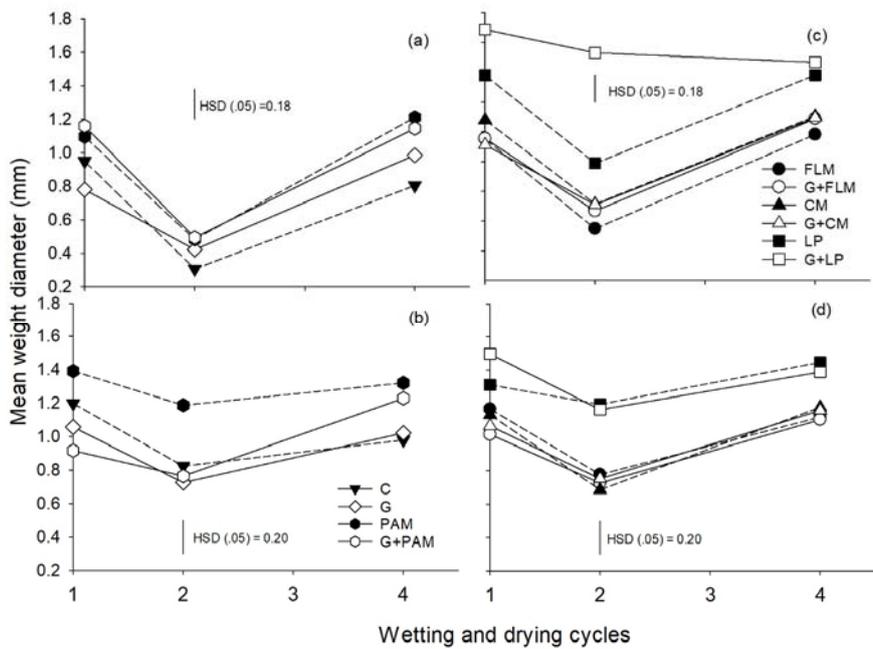
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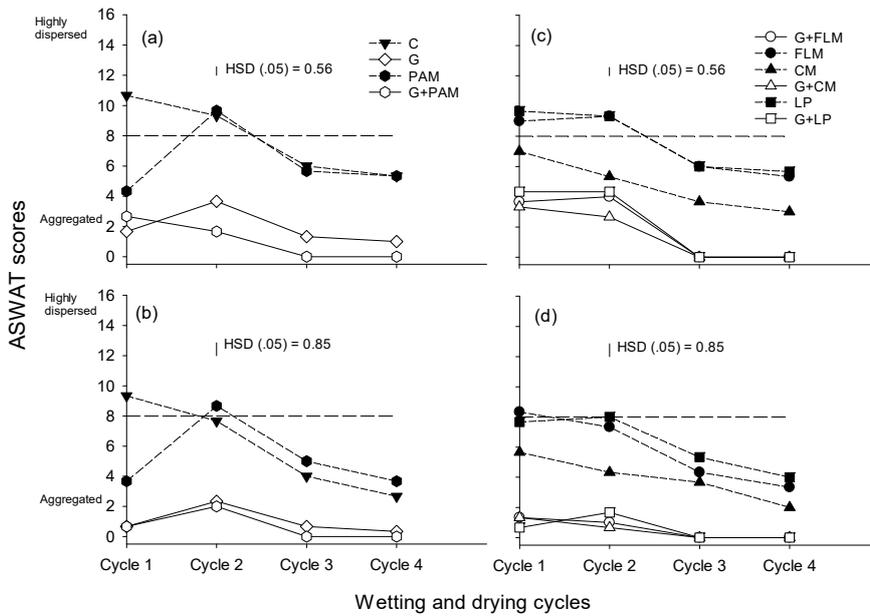
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 386 Figure 2. The mean weight diameter (MWD) of the soils after addition of amendments during four
 387 wetting and drying (WD) cycles. Soil 1 (a, c), and Soil 2 (b, d). The treatments are C: control, G:
 388 gypsum, PAM: anionic polyacrylamide, FLM: feedlot manure, CM: chicken manure, LP: lucerne
 389 pellets. Vertical bars represent Tukey's honest significant difference (HSD) values at $P=0.05$ for
 390 pairwise comparisons among cycles.

391 **3.2 Soil dispersion dynamics as measured by ASWAT and DI**

392 We assessed soil dispersion using both the ASWAT test (Fig. 3) and DI (see Supplementary
 393 Data Fig. S2), with good agreement between the DI results and ASWAT scores (Fig. S3).
 394 Overall, we observed that soil dispersion decreased with WD cycles in both soils. Although,
 395 Soil 1 was more dispersive than Soil 2, both soils showed a similar decrease in dispersion with
 396 WD cycles. Addition of gypsum significantly decreased dispersion in all WD cycles, and this
 397 effect of gypsum was greater than the effect of organic amendments in decreasing soil

398 dispersion. PAM had only a short-term effect, with PAM decreasing dispersion for the first
 399 WD cycle but not thereafter. Of the organic amendments, only CM decreased the dispersion in
 400 both soils, but FLM and LP did not decrease dispersion in both soils in the first and second
 401 WD cycles. Given that both EC (Fig. S4) and SAR (Fig. S5) are known to affect soil dispersion,
 402 it was not surprising that the ASWAT scores were negatively correlated with EC (Fig. S6), and
 403 positively correlated with SAR (Fig. S7). However, the ASWAT scores became less affected
 404 by SAR after second WD cycle, with this highlighting the importance of WD cycles on
 405 structure improvement.

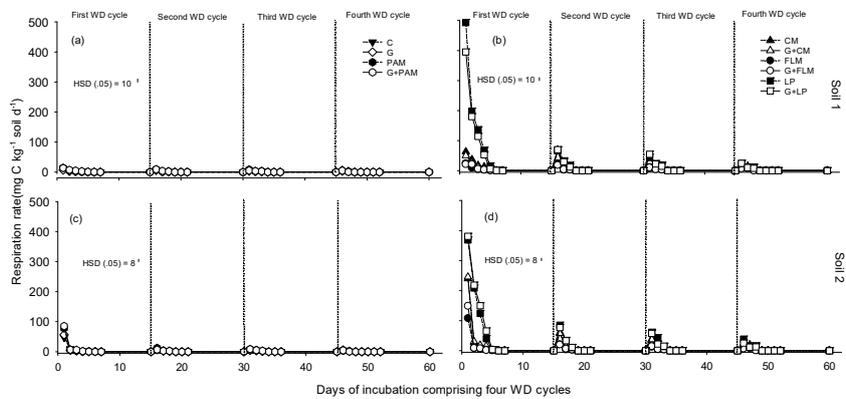


406
 407 *Figure 3. The ASWAT scores of soils after addition of amendments during four wetting and drying*
 408 *(WD) cycles; Soil 1 (a, c), and Soil 2 (b, d). The amendments are C: control, G: gypsum, PAM: anionic*
 409 *polyacrylamide, G+PAM: gypsum+anionic polyacrylamide, FLM: feedlot manure, G+FLM:*
 410 *gypsum+feedlot manure, CM: chicken manure, G+CM: gypsum+chicken manure, LP: lucerne pellets,*

411 *G+LP: gypsum+lucerne pellets*). Vertical bars represent Tukey's honest significant difference (HSD)
412 values at $P=0.05$ for pairwise comparisons among cycles.

413 3.3 Soil microbial respiration

414 Soil WD cycles impose a significant stress on the soil microbial community. The addition of
415 treatments resulted in a significant increase ($p<0.001$) in soil microbial respiration for both the
416 soils (Fig. 4). Rewetting of the dried soils caused a transient increase in microbial respiration
417 (Fig. 4), with respiration being highest during the first WD cycle and decreasing during
418 subsequent WD cycles. The lowest microbial respiration rates were observed in the fourth WD
419 cycle in both soils (Fig. 4). The highest microbial respiration rates were observed in soils
420 amended with LP or CM in the first WD cycle, whereas FLM had little effect on microbial
421 respiration, whilst PAM and gypsum had no effect on microbial respiration.



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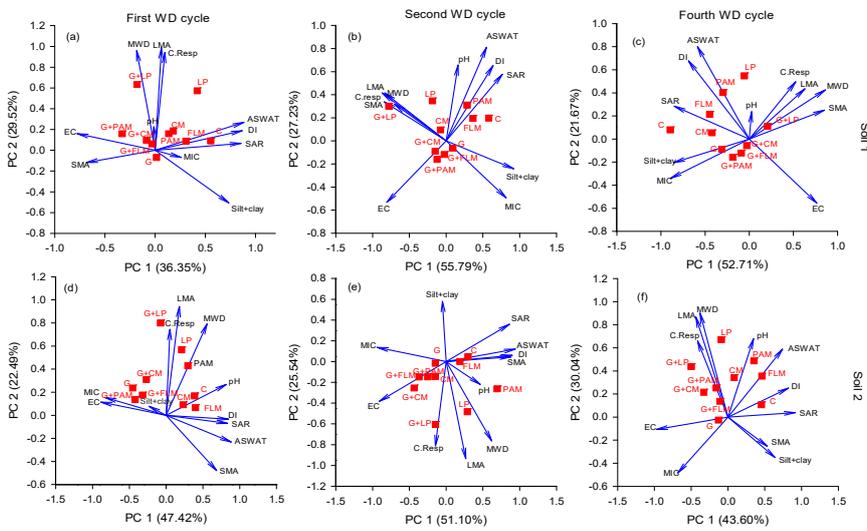
423 *Figure 4. Changes in soil respiration rate after addition of amendments during four alternate WD*
 424 *cycles; Soil 1 (a, b) Soil 2 (c, d) The treatments were C: control, G: gypsum, PAM: anionic*
 425 *polyacrylamide, G+PAM: gypsum+anionic polyacrylamide, FLM: feedlot manure, G+FLM:*
 426 *gypsum+feedlot manure, CM: chicken manure, G+CM: gypsum+chicken manure, LP: lucerne pallets,*
 427 *G+LP: gypsum+lucerne pallets. Vertical bars represent Tukey's honest significant difference (HSD)*
 428 *values at P=0.05 for pairwise comparisons among the four WD cycles. The vertical dotted lines are*
 429 *showing the days when deionised water was added to start the next WD cycle.*

430 3.4 Relationship between soil physical, chemical, and microbial properties

431 Soil properties determined for both soils after the first, second, and fourth WD cycles (Fig. 5)
 432 were used to examine inter-relationships by PCA. Overall, the PCA biplots indicated that soil
 433 microbial respiration, MWD, and the proportion of large macroaggregates were positively
 434 correlated with each other. A positive correlation between ASWAT, DI with SAR was
 435 observed in both soils, whereas EC had a negative correlation with ASWAT and DI.
 436 Surprisingly, there was not a strong correlation between silt+clay and DI or ASWAT scores.
 437 The effect of SAR and EC on soil dispersion decreased after the second WD cycle (Fig. 5),
 438 underlining the importance of WD cycles on structure formation and stability, which offsets
 439 the detrimental effects of SAR on stability. Obvious treatment effects on different soil
 440 properties could also be seen in each WD cycle (Fig. 5). The G+LP and LP treatments [were](#)

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441 positively correlated with microbial respiration, the proportion of large macroaggregates and
442 MWD in all the cycles. The gypsum added with and without organic amendments produced
443 the highest EC in all the cycles. The control soils had highest SAR, ASWAT scores and DI
444 followed by FLM and LP. It was also observed that the proportion of large macroaggregates
445 and the MWD were negatively correlated with the silt+clay fraction in each WD cycle.



446
447 Figure 5. Principal component analysis (PCA) biplot of the physicochemical and microbial properties
448 effected by different treatments in two soils after first, second, and fourth WD cycle: Soil 1(a, b, and c)
449 and Soil 2 (d, e, and f). The treatments are C: control, G: gypsum, PAM: anionic polyacrylamide, FLM:
450 feedlot manure, CM: chicken manure, LP: lucerne pellets. The data presented in these PCA biplots are
451 the mean data of all the days for each parameter (blue arrows) and factorial scores of all the treatments
452 (red squares). The parameters are ASWAT scores (aggregate stability in water test), DI (Dispersion
453 index), C. Resp (cumulative microbial respiration), SAR (sodium adsorption ratio), EC (electrical
454 conductivity 1:5), LMA (large macroaggregates), SMA (small macroaggregates), MIC
455 (microaggregates), Silt+Clay, and MWD (mean weight diameter).

457 **4 Discussion**

458 The results of this experiment showed clear differences in the role of organic amendments,
459 gypsum (Ca), and WD cycles on the formation and stabilisation of soil aggregates. We showed
460 that the formation and stability of large macroaggregates was controlled by the type of organic
461 amendments, whereas the WD cycles increased the formation and stability of small
462 macroaggregates. Formation of microaggregates is beneficial in that it decreases dispersion,
463 but microaggregates may not be sufficient to improve soil water infiltration (Collis-George and
464 Greene 1979; Nemati *et al.* 2002). Regardless, even a small increase either in large
465 macroaggregates and small macroaggregates may increase infiltration, with this facilitating the
466 leaching of excess Na. Here, we discuss the importance of each factor (WD cycles, organic
467 amendments, and Ca) individually.

468 **4.1 The relative role of WD cycles on aggregation**

469 The WD cycles result in rearrangement of pores and soil particles and may lead to increased
470 rigidity and stability of soil aggregates (Horn *et al.* 2014). We observed a marked change in
471 aggregate size distribution with repeated WD cycles (Fig. 1), from macroaggregates (large
472 macroaggregates and small macroaggregates) at the completion of the first WD cycle, to
473 microaggregates at the completion of the second WD cycle, and back to macroaggregates (large
474 macroaggregates and small macroaggregates) at the completion of the fourth WD cycle. We
475 suggest that extracellular polysaccharides formed by microbial activity (indicated by soil
476 microbial respiration, Fig. 3) are responsible for the formation of large macroaggregates at the
477 completion of the first WD cycle. After the second WD cycle, the microbial activity greatly
478 decreased (Fig. 3) and macroaggregates (large macroaggregates and small macroaggregates)
479 were broken down into microaggregates and silt+clay. This allowed the soil particles to settle
480 into tightly packed configurations, resulting in stronger interconnections upon WD cycles
481 (Kemper and Rosenau 1984). Macroaggregates were more susceptible to disintegration during

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486 wet sieving compared to microaggregates at the completion of second WD cycle. By the fourth
487 WD cycle, some rearrangements of soil particles likely occurred, facilitated by soil drying,
488 thereby rebuilding macroaggregates. The decrease in large macroaggregates (Fig.1) at the
489 completion of the first WD cycle is also consistent with the findings of Denef *et al.* (2001).
490 Rahman *et al.* (2018) and Zhang *et al.* (2022). Macroaggregates are expected to be more
491 susceptible to disintegration due to water content changes, because of the large number of
492 planes of weakness and greater angular momentum (Kay, 1990; Denef *et al.* 2001; Zhang *et al.*
493 2022). The changes observed in the proportion of small macroaggregates followed the same pattern
494 as large macroaggregates, with an initial increase in the proportion of small macroaggregates
495 when the soils were treated with organic amendments. In contrast to these results, no significant
496 differences were observed when the same soils were treated with organic amendments under
497 continuous wet conditions (Niaz *et al.*, 2022). Also, the increase in proportion of larger
498 aggregates (large macroaggregates and small macroaggregates) is 2x greater when the same
499 soils were exposed to WD cycles as compared to constant wet regime (Niaz *et al.*, 2022). In
500 this study, the proportion of small macroaggregates did not increase after the second WD cycle,
501 but the proportion of microaggregates and silt+clay fraction increased, suggesting that upon
502 repeated WD cycles, large macroaggregates break down into microaggregates and silt+clay
503 fractions. These observations support the finding that macroaggregates are composed of
504 microaggregates (Six *et al.*, 2000a). In addition, we observed that the control soils (where no
505 amendments were added) also showed an increase in the proportion of small macroaggregates
506 with WD cycles. This highlights the importance of WD cycles and suggests that the formation
507 of small macroaggregates is less dependent on the organic matter content, in agreement with
508 the PCA biplots (Fig. 5). The significant increase in the proportion of small macroaggregates
509 in control soils after the fourth WD cycle also indicates that there could be an increase in

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516 interparticle bond strength with ageing or time (Utomo and Dexter 1982; Dexter 1988; Kong
517 *et al.* 2005; Bravo-Garza *et al.* 2009).

518 4.2 The role of organic amendments on soil respiration and aggregation

519 The addition of organic amendments provided an energy and nutrient source for
520 microorganisms and resulted in increased respiration rates, although we observed that
521 respiration rate decreased with repeated WD cycles (Fig. 4), presumably due to depletion of
522 easily metabolisable organic compounds (Harrison-Kirk *et al.* 2013; Yu *et al.* 2014; Rahman
523 *et al.* 2018; Fraser *et al.* 2016; Brangari *et al.* 2022; Zhang *et al.* 2022). In contrast to LP, CM
524 had a modest effect on microbial respiration, whereas FLM had no significant effect on
525 microbial respiration as it was already decomposed. Rewetting of dry soil led to immediate
526 flush of microbial respiration (Fig. 4) similar to the results of Brangari *et al.* (2022). This could
527 be attributed to the slaking of larger aggregates (Fig. 1), thereby exposing some occluded
528 organic matter, or due to the utilization of substrates that become available upon rewetting (Wu
529 and Brookes 2005; Borken and Matzner 2009; Yu *et al.* 2014; Zhang *et al.* 2022). These
530 available (labile) substrates after rewetting include remnants of the added organic matter and
531 microbial biomass (Wu and Brookes 2005; Zhang *et al.* 2022). Over time, with successive WD
532 cycles, it is likely that the labile organic C pool was either exhausted or became physically
533 protected within aggregates and hence became less accessible to microbial degradation (Zhang
534 *et al.* 2022).

535 For the LP and G+LP treatments, where the increase in respiration rate was ~4-fold greater in
536 Soil 1 and ~2 fold greater in Soil 2 than for any other treatment (Fig. 4), we observed that the
537 soils also had a significant increase in the proportion of large macroaggregates (Fig. 1) and
538 MWD (Fig. 2), suggesting that labile organic compounds are important for structure formation
539 (also confirmed by PCA biplots Fig. 5). The formation of large macroaggregates after
540 incorporation of organic amendments, as observed here for LP, has also been reported by Deneff

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549 *et al.* (2001), Bravo-Garza *et al.* (2009) and Rahman *et al.* (2018). Although the maximum
550 respiration rate was observed during the first WD cycle for LP and G+LP, the proportion of
551 large macroaggregates and MWD increased at the end of fourth WD cycle. During later WD
552 cycles, it is likely conversion of readily-metabolisable organic compounds (e.g., microbial
553 polymers) to more resistant forms resulted in the formation of macroaggregates (large
554 macroaggregates and small macroaggregates), but in this case, aggregation may have been
555 caused by more hydrophobic compounds. However, the proportion of large macroaggregates
556 did not increase much as compared to the first WD cycle, likely because microbial activity was
557 lower (Cosentino *et al.* 2006; Zhang *et al.* 2022). One of the possible reasons of increased
558 MWD could be the accumulation of microbial binding agents over time that are released
559 continuously from microbial activity due to organic matter decomposition (Rahman *et al.*
560 2018).

561 When the soils were treated with organic amendments, a decrease in dispersion was observed,
562 but was not significantly different from control except in CM treated soils. The possible reason
563 for increased stability in CM treated soils was its higher Ca content (Table 2), increased EC
564 (Fig. S4) and decreased SAR (Fig. S5). For the remaining organic amendments viz; FLM, LP,
565 and PAM, an improvement in aggregate stability was observed after the second WD cycle.
566 Although, LP had the highest MWD, it still did not reduce dispersion in the first WD cycle.
567 We suggest that high MWD in itself is a misleading measure of soil stability since only a few
568 large aggregates can result in a high MWD (Niaz *et al.*, 2022). Some dispersion can occur in
569 parallel which is sensitively detected by the ASWAT test, but not by the MWD. Both assays
570 detect different physical processes - hence a high MWD (Fig. 1) and dispersion (Fig. 2) are not
571 mutually exclusive (Niaz *et al.*, 2022, Organic amendments rich in aliphatic compounds or
572 waxes can lead to hydrophobicity of aggregate surfaces during the drying step which slows
573 down the wetting of aggregate surfaces reducing the disruption caused during rewetting

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578 (Piccolo and Mbagwu 1999; Borken and Matzner 2009). Monnier (1965) proposed that fresh
579 organic amendments can increase aggregate stability in the time frame of weeks and months as
580 compared to already decomposed stable organic amendments. But in our study, we found no
581 significant differences in dispersion after the addition of LP (fresh) and FLM (partially
582 decomposed). These findings suggest that there might be some other mechanisms which are
583 responsible for the binding of organic matter with clay particles to control dispersion, such as
584 the size of the organic matter molecule and charge density. This requires further investigation.

585 4.3 The relative role of Ca (gypsum) in improving aggregate stability

586 The results of this experiment showed that addition of gypsum (Ca) significantly reduced soil
587 dispersion and increased aggregate stability. This improvement in aggregate stability is because
588 of the increased EC (ionic strength, Fig. S4) and decreased SAR (Fig. S5) after the addition of
589 gypsum. The increased EC likely resulted in the flocculation of soil particles by reducing the
590 diffuse double layer (van Olphen 1977, Ghosh et al., 2010, Bennett et al. 2015). Improved
591 stability was also observed when organic amendments were applied with gypsum especially in
592 G+PAM, G+LP, and G+FLM treated soils. The PAM had an initial positive effect but led to
593 decreased stability at completion of the second WD cycle. Although the addition of gypsum
594 increased aggregate stability, it was observed that addition of gypsum did not affect the
595 proportion of large macroaggregates and MWD. However, when organic amendments were
596 added with gypsum an improvement in proportion of large macroaggregates and MWD was
597 observed in G+LP treated soils. This can be explained as Ca-bridging effect through which
598 clay particles are attached to organic matter and polyvalent cations, resulting in the formation
599 of macro and micro aggregates (Wuddivira and Camps-Roach 2007).

600 Conclusion

601 The stability of dispersive sodic Vertisols was improved by the application of organic
602 amendments and gypsum, which was further enhanced by WD cycles. Gypsum reduced soil

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Deleted: Aggregate formation in two sodic Vertisols was significantly affected by WD cycles, organic matter amendments and gypsum. However, we observed that not all organic amendments were beneficial for increasing the formation of macroaggregates. Organic amendments LP and CM increased the proportion of larger aggregates, whereas FLM was ineffective which can be attributed to the effects of amendments on microbial activity. While G helped in reducing soil dispersion, it had no significant role in the formation of larger aggregates. Implementation of these findings in the field would favour the use of organic amendments with gypsum to improve the physicochemical properties of sodic soils, which is further enhanced by WD cycles. The aim should be initially to prevent soil dispersion which can be achieved by the application of Ca (through application of gypsum) and then to build larger aggregates which can be achieved by the application of organic amendments, and imposing wetting and drying cycles. The large aggregates then improve water and air entry in soil.¶

641 dispersion but did not affect the proportion of large macroaggregates and MWD. We observed
642 that not all organic amendments were equally beneficial in improving soil aggregation and
643 aggregate stability. LP significantly increased the proportion of large macroaggregates
644 compared to FLM and PAM. In contrast, CM significantly reduced soil dispersion as it had
645 higher calcium content. It was also found that PAM only had a transient effect in controlling
646 dispersion. In the absence of organic amendments, repeated WD cycles reduced the dispersion
647 of sodic soils, but when organic amendments were added (with or without gypsum) soil
648 aggregation and soil stability was improved even more. It is likely that soil microbial activity
649 contributed to the aggregate formation. Implementation of these findings in the field would
650 favour the use of organic amendments with gypsum to improve the physicochemical properties
651 of sodic soils, which is further enhanced by WD cycles. The aim should be initially to prevent
652 soil dispersion which can be achieved by the application of Ca (through application of gypsum)
653 and then to build larger aggregates which can be achieved by the application of organic
654 amendments.

655 **Author contribution**

656 Conceptualization: Niaz, S., Wehr, J. B., Kopittke, P.M., Dalal, R.C., and Menzies, N.W.;;
657 Formal analysis, investigation, and writing original draft: Niaz, S; Review and editing: Wehr,
658 J. B., Kopittke, P.M., Dalal, R.C., and Menzies, N.W.; Supervision: Wehr, J. B. and Menzies,
659 N.W.

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660 **Competing interests**

661 The authors declare that they have no conflict of interest.

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